Asian-Australasian Journal of Bioscience and Biotechnology

ISSN 2414-1283 (Print) 2414-6293 (Online) https://www.ebupress.com/journal/aajbb/

Article

Surveillance and prevalence of gastrointestinal parasite of domestic animals in different abattoirs in Bangladesh

Md. Murshed Hasan Mustafa^{1,2*}, Md. Rafiul Islam¹, Md. Abul Hashem², Md. Abdul Alim³ and Md. Mukhlesur Rahman²

*Corresponding Author: Md. Murshed Hasan Mustafa, Deputy Director (Livestock), Bangabandhu Academy for Poverty Alleviation and Rural Development (BAPARD), Ministry of LGRD and Co-operatives, Gopalganj, Bangladesh. E-mail: murshed9137@yahoo.com

Received: 16 May 2022/Accepted: 27 July 2022/Published: 28 August 2022

Copyright © 2022 Md. Murshed Hasan Mustafa *et al*. This is an open access article distributed under the Creative Commons Attribution 4.0 International License (https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Abstract: The study was conducted to examine the gastro-intestinal parasite infestation of animal at Gopalganj district in Bangladesh during July 2020 to June 2021. Total collected sample were 151, among them 32 cattle and goat 119. Parasites were collected directly from viscera then preserve for further analysis. Age, sex and species of the examined animals were recorded as far as practicable. Overall prevalence of nematodes, cestodes and trematodes were 62.25%, 9.27% and 50.33% respectively. The prevalence of parasites was variable with *Paramphistomum* spp. (58.29%), *Fasciola* spp 62.91%, *Moniezia* spp. (5.3%), *Haemonchus* spp. (41.06%), *Trichuris* spp. (3.31%), *Oesophagostomum* spp. (10.59%), *Strongyloides* spp. (2.64%), *Trichostrongylus* spp. (5.29%), *Bunostomum* spp. (5.96%), Toxocara spp. (25.17%). Parasitic in male and female exhibited no significant variations (p>0.05) between them. The infection was significantly higher in young than in adult. No significant (p>0.05) variation was observed in infection rates between cattle and goats.

Keywords: gastrointestinal parasite; prevalence; cattle; goats; abattoirs; BAPARD

1. Introduction

Bangladesh is an agricultural country where 21% GDP comes from the agricultural sector like crops, livestock, fisheries and forestry (Begum *et al.*, 2007; Hasan *et al.*, 2021; Mustafa *et al.*, 2020 and 2021). There are about 24.54 million cattle, 1.5 million buffalo, 3.68 million sheep and 26.6 million goat and also share in agricultural GDP 13.1%. (Livestock Economy, 2020-21). Livestock demand due to low nutritional status and disease prevalence (Hasan *et al.*, 2022; Barman *et al.*, 2017; Sarkar *et al.*, 2008; Mazed *et al.*, 2004; Rahman *et al.*, 2002, 1999, 1998 and 1997). Parasitism claims to be one of the main obstacles in livestock production in Bangladesh (Islam *et al.*, 2022; Mustafa *et al.*, 2022; Jabbar *et al.*, 1983; Khokan *et al.*, 2017). Gastrointestinal parasitism is a world-wide problem and in Bangladesh it may be considered as one of the major constraints in cattle production. The infection causes productivity losses through reduced feed intake and decreased efficiency in feed utilization due to subclinical or chronic infections that are responsible for economic losses (Renaldi *et al.*, 2011; Sarker *et al.*, 2015; Bary *et al.*, 2018).

¹Bangabandhu Academy for Poverty Alleviation and Rural Development (BAPARD), Ministry of LGRD and Co-operatives, Gopalganj, Bangladesh

²Department of Animal Science, Bangladesh Agricultural University, Mymensingh-2202, Bangladesh

³Department of Parasitology, Bangladesh Agricultural University, Mymensingh-2202, Bangladesh

Gastrointestinal parasites like helminths and protozoan are very common in cattle and goats. When heavy infections occur those parasites contribute to reduced milk and meat production (Murthy and Rao, 2014). The losses caused by parasitic infections are in the form of lowered general health condition, retarded growth rate, diminishing the working efficiency, decrease milk and meat production, abortion; cost associated with preventive measures and reduces the disease resistance capability, which may ultimately lead to higher mortality (Silvestre et al., 2000). Small ruminants under intensive and extensive production systems are extremely susceptible to the effects of wide range of helminths (Abede and Esayas, 2001). Helminth infestation lowers the immunity of the animals and render them susceptible to other pathogenic infections (Garedaghi et al., 2011: Sarker et al., 2020). Rahman et al. (1975) reported the death of as high as 25.0% kids and lambs and 43.5% adult goats due to GI parasites in both rural and farm condition. In another report, Mazid et al. (2006) reported that about 81.1% and 94.7% helminthiasis occurred in cattle and goats, respectively. Various risk factors related to host and environment play an important role in the onset of GI parasitic infections. Environmental factors include agro-ecological conditions, animal husbandry practices such as housing system, deworming intervals and pasture management; these largely determine the type, incidence and severity of various parasitic diseases (Badran et al., 2012; Mustafa, 2022). Parasitic problems are often neglected and overlooked as majority of the infected animals show a number of little obvious clinical signs during their productive life and their effects are gradual and chronic (Islam et al., 1985; Tariq et al., 2010).

Gopalganj district is very prospective for cattle and goat rearing due to its agro-ecological condition. Despite routine vaccination against major infectious diseases, large and small ruminants are still suffering from poor body condition state due to parasitism. However, there is limited information about the infection prevalence of GI parasites in large and small ruminants in the study area. Therefore, the present study was conducted to investigate the GI parasites prevalent in Gopalganj district and identify associated risk factors such as age, sex and species.

2. Materials and Methods

2.1. Study site and study period

The study areas were the surroundings of Gopalganj district, Bangladesh. Samples were collected during the period from July 2020 to June 2021. A total of 151 carcasses of cattle and goats were examined for GI parasites in different abattoirs in Gopalganj region of Bangladesh.

2.2. Sample collection and preservation

A total of 151 fecal samples were collected from different abattoirs of Gopalganj region irrespective of age, sex, breed, BCS (body condition scored) and seasons like rainy, summer and winter. In this study, age of cattle are categorized into three groups, viz calf aged 3 less than, aged > 3years, goat \le 2 years and adult aged > 2 and above. During sample collection, the biometric data of each animal like, age, sex breed, and general body condition were recorded. Approximately 10 gm of fecal samples were collected directly from the rectum of the cattle and kept into a small plastic container and stored in ice containing cool box and labeled properly. After collection, the fecal samples were preserved with 10% formalin and transferred immediately to the Laboratory of the BAPARD and stored at 4°C temperature until further examination.

The sample for *Fasciola* spp., *Paramphistomum* spp. were collected in normal saline and washed in distilled water through several changes and kept in glycerinealcohol (70% ethyl-alcohol 95 parts with pure glycerine 5 parts). Helminth parasites (*Fasciola* spp.) in the liver were collected and examined in fresh state. They were shaken vigorously in 1 percent salt solution and then fixed in 10 percent formalin. This was done to prevent contraction of other parasites. Sometimes a fixative consisting of 85 parts of 85% ethyl-alcohol, 10 parts of 40% formalin and 5 parts of glacial acetic (AFA) was used to fix the parasites.

Parasites were also collected from different parts of the body of the individual cattle by hand picking and precautions were taken to preserve the mouthparts and appendages of the GI parasites during collection. Parasites were preserved in 70% alcohol in a clean and well stopper glass vials which were labeled properly.

2.3. Laboratory examination of fecal samples and parasites

Fecal samples were examined using the direct smear method followed by the McMaster technique and eggs were identified under microscope at low power objective (10X) followed by higher magnification (40X objectives) according to the technique by Urquhart *et al.*, 1996.

The positive samples in the direct smear method were subjected to EPG counting to determine the number of eggs per gram of feces. Eggs were identified according to the previously described morphological characteristic by Soulsby (Soulsby, 1982). The EPG (egg per gram of feces) were calculated by multiplying the total numbers

of identified eggs in two chambers with the dilution factor 50. The parasitic load was considered as light (egg counts from 1 to 500), moderate (egg counts above 500 to 1000) and heavy or severe infection respectively when eggs counts were found >1000 per gram of feces and parasites were identified under light microscopy based on their morphological characters (Sweeny *et al.*, 2011).

2.4. Statistical analyses

Statistical analyses were carried out by Statistical Package for Social Science (SPSS version 22.0, SPSS Inc., Illinois, USA) using F test. To identify the risk factors univariate analysis was performed.

3. Results and Discussion

3.1. Overall prevalence of gastrointestinal parasitism

In this study a total of 151 carcasses of cattle and goat (32 cattle and 119 Goat sample) were examined at different abattoirs of Gopalganj region. Overall prevalence of nematodes, cestodes and trematodes was 62.25%, 9.27% and 50.33% respectively (Table 1). The prevalence of parasites were varied with *Fasciola* spp. (62.91%), *Paramphistomum* spp. (58.29%), *Moniezia* spp. (5.3%), *Haemonchus* spp. (41.06%), *Trichuris* spp. (3.31%), *Oesophagostomum* spp. (10.59%), *Strongyloides* spp. (2.64%), *Trichostrongylus* spp. (5.29%), *Bunostomum* spp. (5.96%), Toxocara spp. (25.17%) (Table 2).

Table 1. Prevalence of parasites in slaughtered animals.

Parasite	Cattle (N=32)	Goat (N=119)	Total (N=151)
Nematodes	62.5% (20)	62.18% (74)	62.25% (94)
Cestodes	9.37% (3)	9.24% (11)	9.27% (14)
Trematodes	46.87% (15)	51.26% (61)	50.33% (76)

Table 2. Prevalence of different parasites in slaughtered animals.

Parasite	Cattle (N=32)	Goat (N=119)	Total (N=151)
Fasciola spp.	68.75% (22)	61.34% (73)	62.91% (95)
Paramphistomum spp.	53.12% (17)	59.66% (71)	58.29% (88)
Moniezia spp.	3.12% (1)	5.88% (7)	5.3% (8)
Haemonchus spp.	37.5% (12)	42.02% (50)	41.06% (62)
Trichuris spp.	3.12% (1)	03.36% (4)	03.31% (5)
Oesophagostomum spp.	6.25% (2)	10.08% (12)	10.59% (16)
Strongyloides spp.	6.25% (2)	01.68% (2)	2.64% (4)
Trichostrongylus spp.	09.37% (3)	04.20% (5)	05.29% (8)
Toxocara spp.	34.37% (11)	22.69% (27)	25.17% (38)
Bunostomum spp.	0%	05.96% (9)	05.96% (9)

The same findings was partially consistence with the reports of Khan *et al.* (2010) and Saravanana *et al.* (2009) who documented slightly lower prevalence in Pakistan and India, respectively. The reports of Hirani *et al.* (2006) supported the prevalence of Trematode (46.25%) of the present study. The earlier findings of indigenous cattle was also partially similar with the findings of Raza *et al.* (2007) and Regassa *et al.* (2006) who documented somewhat higher prevalence in Pakistan and Ethiopia, respectively. However, similar types of parasites as reported in this study were detected by different scientists in different areas with variable rate of infections (Ahmed *et al.*, 2015; Islam *et al.*, 2014; Xiao, 2010; Mustafa *et al.*, 2022). Variation in the occurrence of gastrointestinal parasitic infections might be due to geo-climatic conditions, breed, age, sex, plane of nutrition, stress, availability of intermediate host (Hansen and Perry, 1993).

3.2. Sex-specific prevalence of gastrointestinal parasitism

The prevalence of livestock parasite in cattle and goat is variable in relation to sex. In this study, Livestock parasite present higher in Bulls (66.67%) than cows (50%) but in case of goat, doe bears higher infection than buck and castrated goat (Table 3).

Table 3. Prevalence of parasitic diseases in several species in relation to sex.

Parasite	Catt	Cattle (N=32) Goat (N=119)			
	Male	Female	Male	Female	
	N=24	N=08	N=41	N=78	
Nematodes	66.67% (16)	50% (4)	56.09% (23)	65.38% (51)	
Cestodes	12.5% (3)	0%	07.32% (3)	10.26% (8)	
Trematodes	41.67% (10)	62.5% (5)	41.46% (17)	56.41% (44)	

Sex-specific prevalence of gastrointestinal parasitic infections showed that infection caused by Nematode, Cestode and Trematode were found predominant in female than male cattle. Findings of this study was found in accordance with the reports of Raza *et al.* (2007, 2010) who also reported higher worm burden in female cattle compared to male cattle. On other hand, Nematode infection in indigenous male cattle was found in harmony with the reports of Rekwot and Ogunsusi (1985) and Soulsby (1982). In crossbred cattle, Nematode infections were also in line with the observation of Avcioglu and Balkaya, (2011) and Raza *et al.* (2010). Variation in occurrence of such helminth infections in male and female animals might be due to the variation in sample size (Bachal *et al.*, 2002), lowered resistance of female animals or temporary loss of acquired immunity near parturition (Garcia *et al.*, 2007), stress, genetic resistance of host and insufficient feed supply against their higher needs (Raza *et al.*, 2010; Hansen and Perry, 1993).

3.3. Age specific prevalence of gastrointestinal parasitism

The prevalence of gastrointestinal parasitic infections varied between two age groups of cattle. The highest infection of Nematode, Cestode and Trematode was recorded in cattle of >3 years (54.55%, 0%, 54.55% respectively).

Table 4. Prevalence of parasitic diseases in relation to age of cattle.

Types of Parasites	Age			
	< 3 year N=21	Prevalence	>3Year N=11	Prevalence
Nematodes	14	66.67%	06	54.55%
Cestodes	03	14.29%	00	0%
Trematodes	09	42.86%	06	54.55%

Age specific prevalence (Table 4) of gastrointestinal parasitic infections especially, Nematode, Cestode and Trematode were found more in adult cattle which supported the observation of Sardar *et al.* (2006) who reported that Nematode, Cestode and Trematode were highest in the age group greater than 36 months and lowest in age group less than 12 months. Findings of Fritsche *et al.* (1993) also supported the findings of this study. The earlier findings of this investigation showed disagreement with Raza *et al.* (2007) and Regassa *et al.* (2006) who recorded significantly higher worm burden in younger animals than adult. Higher prevalence of parasitic infection in adult cattle might be due to keeping them for a longer period of time in breeding and milk production purposes or supply inadequate feed against their high demand (Sardar *et al.*, 2006). Moreover, stress like lactation, pregnancy, nutritional deficiency which might be accounted for higher prevalence in adult cattle (Radostits, 1994). On the other hand, the highest prevalence of Nematode infection in calf was supported by the reports of Sarder *et al.* (2006) and Bachal *et al.* (2002) who recorded such infection in early months of life.

4. Conclusions

This surveillance work gives an outline about GI parasitic infection cattle and goat in Gopalganj area in Bangladesh that will be helpful to design a layout for controlling parasitic infestation, training manual preparation of BAPARD and further action research about parasitic infection in this area.

Acknowledgements

The authors gratefully acknowledge the Bangabandhu Academy for Poverty Alleviation and Rural Development (BAPARD), Ministry of LGRD and Co-operatives, Gopalganj, Bangladesh for funding the research works. The authors also acknowledge the Department of Animal Science and the Department of Parasitology, Bangladesh Agricultural University, Mymensingh for their technical assistance.

Data availability

The data used to support the findings of this study are included within the article.

Conflict of interest

None to declare.

Authors' contribution

Md. Murshed Hasan Mustafa wrote the manuscript, Md. Rafiul Islam collected samples from abattoirs and examined them, Md. Abul Hashem assisted data processing and analyzing, Md. Abdul Alim ckecked the parasitic experimental protocal and Md. Mukhlesur Rahman assisted the manuscript writing and correction of the paper. All authors have read and approved the final manuscript.

References

- Abede W and G Esayas, 2001. Survey of ovine and caprine gastrointestinal helminthosis in eastern part of Ethiopia during the dry season of the year. Rev. de Med. Vet., 152: 379–384.
- Ahmed R, PK Biswas, M Barua, MA Alim, K Islam and MZ Islam, 2015. Prevalence of gastrointestinal parasitism of cattle in Banskhali upazilla, Chittagong, Bangladesh. J. Adv. Vet. Anim. Res., 2: 484-488.
- Avcioglu H and I Balakaya, 2011. Prevalence of *Toxocara vitulorum* in Calves in Erzurum, Turkey. Kafkas. Univ. Vet. Fak. Derg., 17: 345-347.
- Asian Development Bank (ADB), 1984. Asian Development Bank (ADB). 1984. Asian development bank yearbook.
- Bachal B, MS Phullan, R Rind and AH Soomro, 2002. Prevalence of Gastrointestinal Helminths in Buffalo Calves. Biol. Sci., 2: 43-45.
- Badran I, R Abuamsha, R Aref, W Alqisi and J Alumor, 2012. Prevalence and diversity of gastrointestinal parasites in small ruminants under two different rearing systems in Jenin district of Palestine. An-Najah Univ. J. Res., 26: 1–18.
- Bary MA, MZ Ali, S Chowdhury, A Mannan, MNE Azam, MM Moula, ZA Bhuiyan MTW Shaon and MA Hossain, 2018. Prevalence and Molecular Identification of Haemoprotozoan Diseases of Cattle in Bangladesh. Adv. anim. vet., 6: 176-182.
- Begum MAA, MM Hossain, M Khan, MM Rahman and SME Rahman 2007. Cattle fattening practices of selected farmers in Panchagarh district. Bang. J. Anim. Sci., 36: 62-72.
- Blood, D.C. and Radostits, O.M. 1989. Veterinary Medicine .7th edn. Balliere and Tindal, London.
- DLS, 2021. Department of Livestock Services. Livestock Population Report, 2021. Information website: www.dls.gov.bd
- Fritsche T, J Kaufmann and K Pfister, 1993. Parasite spectrum and seasonal epidemiology of gastrointestinal nematodes of small ruminants in the Gambia. Vet Parasitol., 49: 271-283.
- Garcia JA, JG Rodriguez-Diego, G Torres-Hernandez, M Mahieu, EG Garcia, and R Gonzalez-Garduno, 2007. The epizootiology of ovine gastrointestinal strongyles in province of Matanzas. Small Rumin. Res., 72, 119-126.
- Garedaghi Y, AP Rezaii-Saber, A Naghizadeh and M Nazeri, 2011. Survey on prevalence of sheep and goats lungworms in Tabriz abattoir, Iran. J Anim Vet Adv., 5: 1460 1461.
- Hirani ND, JB Solanki, AI Patel, JJ Hasanani, RS Joshi and FP Savaliya, 2006. Prevalence of gastrointestinal parasite in cows of Panjarapols in middle Gujarat. Ind. J. Field Vet., 1: 15-18.
- Hansen J and B Perry, 1993. The Epidemiology, Diagnosis and Control of Helminth Parasites of Ruminants. 2nd Dd. Nairobi, Kenya ILRAD, 20-22.
- Hasan MM, MA Hashem, MAK Azad, MM Billah and MM Rahman, 2022. Fattening practices of beef cattle for quality meat production at Rangpur district of Bangladesh. Meat Research, 2: 15.
- Hasan MM, SME Rahman, MA Hashem, MAK Azad, MR Haque and MM Rahman, 2021. Socio economic status of beef fattened farmers of Kaunia upazila at Rangpur district of Bangladesh. J. Agric. Food Environ., 2: 38-44.
- Islam MR, MMH Mustafa, MM Rahman and MA Alim, 2022. Study on the parasitic infestations in calves at BAPARD surroundings coastal south region in Bangladesh. International Journal of Scientific Engineering and Applied Science, 8: 264-275.
- Islam MR, MG Yasin, MA Al Noman, N Begum and MMH Mondal. 2014. Prevalence of gastrointestinal parasites in cattle at Vangura upazila in Pabna district of Bangladesh. Int. J. Nat. Soc. Sci. 1: 45-52.

- Islam KS, 1985. Present situation of livestock and poultry diseases in Bangladesh. In: Jabbar MA (ed) Bangladesh poshusampadunnayun- Neeti O Kowshal BARC, Dhaka and ADC NY: 84-128.
- Jabbar M and DAG Green, 1983. The status and potential of livestock within the context of agricultural development policy in Bangladesh, The University of Wales, Aberyswyth, UK, p. 113.
- Khan MN, MS Sajid, MK Khan, Z Iqbal and A Hussain, 2010. Gastrointestinal helminthiasis: prevalence and associated determinants in domestic ruminants of district Toba Tek Singh, Punjab, Pak. Parasitol. Res., 107: 787-794.
- Livestock Economy, 2020-21. Department of Livestock Services (DLS), Livestock Economy Report, Bangladesh. www.dls.gov.bd
- Mazed MA, MS Islam, MM Rahman, MA Islam and MA Kadir, 2004. Effect of urea molasses multinutrient block on the reproductive performance of indigenous cows under the village condition of Bangladesh. Pak. J. Biol. Sci., 7: 1257-1261.
- Mazid MA, J Bhattacharjee, N Begum and MH Rahman, 2006. Helminth parasites of the digestive system of sheep in Mymensingh, Bangladesh. Bangladesh J. Vet. Med., 4: 117–122.
- Murthy G and P Rao, 2014. Prevalence of gastro intestinal parasites in ruminants and poultry in Telangana region of Andhra Pradesh. J Parasit Dis., 38: 190–192.
- Mustafa MMH, MR Islam and MM Rahman, 2022. Epidemiological Investigation of Gastrointestinal (GI) Parasite at BAPARD Cattle Farm, Gopalganj in Bangladesh. International Journal of Rural Development, Environment and Health Research, 6: 8-16.
- Mustafa MMH, 2022. A Study on the Role of Women and Men in the Family -BAPARD perspective. International Journal of Academic Multidisciplinary Research, 6: 303-312.
- Mustafa MMH, MR Islam, Hossain MA 2022. Epidemiological Investigation of Paramphistomiasis in Cattle at Kotalipara Upazila Gopalganj District. International Journal of Modern Pharmaceutical Research, 6:17-20.
- Mustafa MMH, MR Islam and MM Rahman, 2021. Effect of ration on growth and cost of production during fattening of upgraded Shahiwal bulls. J. Agric. Food Environ., 2: 44-48.
- Mustafa MMH, MR Islam and MM Rahman, 2020. Effect of feed on the performance of upgraded Holstein Friesian bulls during fattening at BAPARD cattle farm in Bangladesh. Asian J. Med. Biol. Res., 6: 761-767.
- Radostits O, DC Blood and CC Gay, 1994. Veterinary Medicine: A text book of disease of cattle, sheep, pigs, goats and horse. 8th ed. Baillere Tindall Publication, London, 1223- 1225, 1237-1238
- Rahman MM, S Habib, MA Kadir, MN Islam and MS Islam, 2002. Participation of rural people in cattle management practices in a selected area of Bangladesh. J. Anim. Vet. Advances, 1: 196-199.
- Rahman MM, S Akhter, MS Rabbani and MM Hossain, 1999. Indigenous knowledge on livestock practiced by the farmers in Mymensingh district of Bangladesh. Bangladesh J. of Anim. Sci., 28: 97-103.
- Rahman MM, S Akther and MM Hossain, 1998. The availability of the livestock feeds and feeding practices followed by the farmers of some areas of Mymensingh District. Bangladesh J. of Anim. Sci., 27: 119-126.
- Rahman MM, S Akther and MM Hossain, 1997. Socio Economic Aspects of the farmers for livestock keeping in Mymensingh town adjacent areas. Progress. Agric., 8: 153-157.
- Rahman A, 1975. Studies in the diseases of goats in Bangladesh. Mortality of goats under farm and rural conditions. Trop. Anim. Health Prod., 8: 90.
- Rahman MA, MA Islam, AK Talukder, MS Parvin and MT Islam, 2012. Clinical disease of ruminant recorded at the Patuakhali Science and Technology University Veterinary clinic. Bangl. J. Vet. Med., 10: 63-73.
- Raza AM, Z Iqbal, A Jabbar and M Yaseen, 2007. Point prevalence of gastrointestinal helminthiasis in ruminants in southern Punjab, Pakistan. Cambridge University Press, J. Helminthol., 81: 323-328.
- Raza AM, S Murtaza, HA Bachaya, A Qayyum and MA Zaman, 2010. Point Prevalence of Toxocaravitulorum in Large Ruminants Slaughtered at Multan Abattoir. Pak. Vet. J., 30: 242-244.
- Regassa F, T Sori, R Dhuguma and Y Kiros, 2006. Epidemiology of Gastrointestinal Parasites of Ruminants in Western Oromia, Ethiopia Int. J. Appl. Res. Vet. Med., 4: 51-57.
- Rekwot PJ and RA Ogunsusi, 1985. Prevalence of *Toxocara (Neoascaris) vitulorum* infection in cattle around Zairia, Nigeria. J. Anim. Prod. Res., 5: 201-207.
- Saravanan S, AM Dinakaran, J Muralidharan, M Geetha and G Selvaraju, 2009. Prevalence of sub-clinical gastrointestinal parasitic infection in dairy animals. Ind. J. Field Vet., 5: 45-46.
- Sardar SA, MA Ehsan, AKMM Anower, MM Rahman and MA Islam, 2006. Incidence of liver flukes and gastro-intestinal parasites in cattle. Bangl. J. Vet. Med., 4: 39-42.
- Sarkar MM, Hossain MM, Rahman MM and Rahman SME, 2008. Effect of feeding urea molasses block on the productive and reproductive performances of Black Bengal doe. Bangladesh Journal of Bangladesh Agricultural University, 6: 39-46.

- Sarker YA, AH Miah, N Sharif, MH Himel, S Islam, RC Ray, TK Paul, MT Islam and MH Sikder, 2015. A retrospective study of common diseases at veterinary teachinghospital, Bangladesh Agricultural University, Mymensingh. Bangl. J. Vet. Med., 13: 55–61.
- Sarker YA, SZ Rashid, S Sachi, J Ferdous, BL Das Chowdhury, SS Tarannum and MH Sikder, 2020. Exposure pathways and ecological risk assessment of common veterinary antibiotics in the environment through poultry litter in Bangladesh. J. Environ. Sci. Health B, 55: 1061-1068.
- Soulsby EJL, 1982. Helminths, Arthropods and Protozoa of Domesticated Animals, 7th edn. Baillere Tindall, London. 729-735.
- Silvestre A, C Chartier, C Sauve and J Cabaret, 2000. Relationship between helminth species diversity, intensity of infection and breeding management in dairy goats. Vet. Parasitol., 94: 91-105.
- Sweeny JPA, ID Robertson, UM Ryan, C Jacobson and RG Woodgate, 2011. Comparison of molecular and McMaster microscopy techniques to confirm the presence of naturally acquired strongylid nematode infections in sheep. Mol. Biochem. Parasitol., 180: 62-67.
- Tariq K, M Chishti and F Ahmad, 2010. Gastrointestinal nematode infections in goats relative to season, host, sex and age from the Kashmir valley. Indian J. Helminthol., 84: 93–97.
- Urquhart GM, J Armour, JL Duncan, AM Dunn and FW Gennings, 1996. Veterinary Parasitology 2nd edn. Blackwell Science Ltd. UK, 170-176.
- Xiao L, 2010. Molecular epidemiology of cryptosporidiosis: an update. Exp. Parasitol., 124: 80-89.